

Comment

Does the Stress of Laboratory Life and Experimentation on Animals Adversely Affect Research Data? A Critical Review

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Summary — Recurrent acute and/or chronic stress can affect all vertebrate species, and can have serious consequences. It is increasingly and widely appreciated that laboratory animals experience significant and repeated stress, which is unavoidable and is caused by many aspects of laboratory life, such as captivity, transport, noise, handling, restraint and other procedures, as well as the experimental procedures applied to them. Such stress is difficult to mitigate, and lack of significant desensitisation/habituation can result in considerable psychological and physiological welfare problems, which are mediated by the activation of various neuroendocrine networks that have numerous and pervasive effects. Psychological damage can be reflected in stereotypical behaviours, including repetitive pacing and circling, and even self-harm. Physical consequences include adverse effects on immune function, inflammatory responses, metabolism, and disease susceptibility and progression. Further, some of these effects are epigenetic, and are therefore potentially transgenerational: the biology of animals whose parents/grandparents were wild-caught and/or have experienced chronic stress in laboratories could be altered, as compared to free-living individuals. It is argued that these effects must have consequences for the reliability of experimental data and their extrapolation to humans, and this may not be recognised sufficiently among those who use animals in experiments.

Key words: *animal welfare, cost-benefit analysis, data accuracy, glucocorticoids, psychological, stress, translational medical research.*

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Introduction

In the December 2017 issue of ATLA, I published an Editorial, in which I suggested that stress in animals in laboratories, resulting from their environment and experimental procedures, was not limited to welfare concerns (1). It also adversely affects, significantly and unavoidably, multiple biological systems and therefore affects the resultant experimental data. This exacerbates inter-species differences and makes extrapolation to humans even more difficult and unreliable. In an effort to encourage and promote discussion of this issue, which is underappreciated, I am expanding on that Editorial.

Stress may be thought of as “the sum of the biological reactions to any adverse stimulus, physical, mental, or emotional, internal or external, that tends to disturb the homeostasis of an organism” (2). It is increasingly acknowledged that diverse species experience pain, stress and distress (see below), and experience depression and anxiety disorders (see Ferdowsian *et al.* [3]). This includes fish — notably

the ubiquitous laboratory zebrafish — which show emotional fever/stress-induced hyperthermia in response to a variety of stressors, including simple handling (4, 5). Stress in fish can result in increased aggressive behaviour (6), elevated anxiety, diminished weight-gain, and altered levels of dopamine and serotonin metabolites in the brain (7), as well as increased brain levels of extracellular adenosine, which has neuromodulatory effects (8).

While all species experience stressors and stress in their natural environments, psychological and physiological problems can arise with exposure to recurrent stressors, and/or when stress becomes chronic and too difficult to cope with. This leads to allostatic overload (excessive wear and tear on the body), which may manifest in altered physiological responses, some of which can be harmful (9) — generally thought of as a state of distress. However, the US National Research Council (NRC) accepts that there is “confusion in the scientific, regulatory, and animal welfare communities” concerning the distinction between stress and distress; and that these terms are often used interchangeably in animal wel-

fare literature and that relevant available information is “far from complete”, with distress remaining “a complex and still poorly understood phenomenon” (10). Nevertheless, it is generally understood that distress manifests when an individual is unable to cope and adapt successfully to one or more stressors, resulting in compromised well-being due to the inability to return to physiological and psychological homeostasis (11). Notably, an individual can remain in distress, even when a stressor is removed.

While different species and individuals have different stressors, variable ranges of stress to which they can adapt, diverse spectra of tolerance, and dissimilar manifestations and sequelae of excessive stress, they share the same biological pathways and mechanisms that are adversely affected by stress. Briefly, stressors stimulate the hypothalamic–pituitary–adrenal (HPA) axis, the sympathetic adrenal medullary axis, and the sympathetic and parasympathetic nerve projections that directly innervate secondary lymphoid organs (12–14). Among other things, this results in the elevation of the ‘stress hormones’, cortisol and corticosterone (CORT). Raised glucocorticoid (GC) levels, alongside significant elevations of heart rate, blood pressure and other hormone levels, are acknowledged indicators of fear, stress and distress, and are used as biomarkers for stress in terrestrial vertebrates in laboratories (e.g. 15–17).

Direct consequences of these neuroendocrine changes include deleterious effects on innate and adaptive immunity, central nervous system pathology, and cardiovascular and reproductive perturbations (18, 19), leading to extensive and diverse adverse health effects. Psychological trauma, for instance, may result in altered health or damaging patterns of behaviour. In humans, immune perturbations manifest in poor responses to vaccines, increased susceptibility to infections, and accelerated disease progression (see Gurfein *et al.* [18]). Further, the various mechanisms underlying such effects mean that stress has ramifications beyond the individuals experiencing it. Successive generations, and individuals who have experienced prenatal and/or early-life stress, may be destined to suffer the consequences in adulthood.

Caveats and Framing the Argument

Chronic or long-term stress is not unique to animals in laboratories — it is part and parcel of life for all species in whatever environment. I do not intend to suggest otherwise, and it is accepted that animals in the wild experience acute stressors regularly (such as lack of food and shelter, predation, disease, and so on); not only is this entirely normal, but it can be beneficial in terms of building allostatic capacity. This Comment, however, argues

that stresses resulting from life in the laboratory often have negative consequences — given the opportunity, the animals would avoid many laboratory stressors, and so they can hardly be considered benign. In addition, the degree, type, frequency and duration of laboratory stressors are different to those in the wild, and together could be more chronic. This may result in greater adverse consequences for animal welfare and for scientific data quality. Experiencing repeated, unnatural stressors, such as blood draws and gavage, cannot be compared to the brief elevations in CORT resulting from the natural diurnal/nocturnal adrenal cycle. For instance, the former stressors are very different from simply waking up, or being hungry. In well-designed experiments, increases in stress biomarkers are relative to baseline levels in any case (see, for example, 16, 20).

That laboratory stress can — arguably, frequently and unavoidably — have effects that compromise animal welfare and experimental results is accepted by the US NRC’s Committee on Recognition and Alleviation of Distress in Laboratory Animals (10): “In the longer term... a breakdown in an animal’s ability to cope with its environment is likely to lead to adverse emotional states and poor welfare. Some of these cases may be quite minor and not give rise to significant ethical concerns; but prolonged or intense circumstances would compromise the animal’s welfare enough to warrant concern and also significantly affect the research results.” Further: “Strong evidence in rodents has shown that mild stress of 2–3 months duration — a regimen that produces no signs of overt distress — alters the animals’ performance in tests of anxiety, depression, and memory... Other findings indicate that rats’ habituation to a test environment can dramatically affect their response to a toxic substance”. Stress can also be beneficial: “Over a longer time frame, glucocorticoid production in response to infection helps restrict the immune system, thus preventing deleterious effects of inflammatory factors on tissues” (10). While this latter statement is true, it also supports the argument made here, that stress — even when not ‘bad’ — adversely affects biological systems, and consequently confounds research data.

Therefore, it is not only distress that leads to biological consequences affecting data reliability, quality and relevance. It seems clear that the consequences of stress are also an issue for welfare and data quality. To quote again the US NRC: “The impact of distress on both animal welfare and research results is likely even more pronounced than that of stress. Animals exposed to prolonged severe stress experience underlying changes in physiological functions (e.g. gastric lesions or immunosuppression) that can interfere with experimental manipulations; alter experimental variables such as behaviour, drug dosing and clearance; change the progress of a disease; and

contribute to morbidity and mortality. A variety of stressors can contribute to unintended distress, from postoperative pain or infection to barren housing conditions or the solitary confinement of an individual of a social species. Stereotypies, abnormal repetitive behaviours indicative of poor well-being that are often observed in distressed animals, are thought to reflect defective brain function and to be a result of poor animal welfare. Stereotypies are thus likely to interfere with behavioural, neuroscience, and pharmacological studies" (10); and "The impact of stress and distress on the quality of scientific research can result in the generation of compromised data, which in turn necessitates the use of more animals... Both stress and distress represent potential complications in a wide range of experiments, and should be proactively addressed by good experimental design" (10). While I (and others) disagree that the use of more animals and the careful design of experiments can have an acceptably positive impact on welfare, and on the human-relevance of the data generated, the main points in this Comment stand. Efforts to address this issue over many years are acknowledged, but I argue that, due to fundamental biology and inter-species differences, the improvement of husbandry, veterinary care, regulation, oversight, training, etc. can never be sufficient to significantly address and overcome these issues.

Stress Resulting from Laboratory Life and Research, and its Biological Impact

The issue for animals in laboratories is that life can be inherently and excessively stressful — perhaps much more than in their natural environments, from which laboratory conditions differ greatly, even when enriched and accounting for efforts to mitigate stressors. Laboratory conditions preclude or limit many natural behaviours, housing tends to be much smaller than the animals' natural ranges, and the animals are subjected to frequent manipulations and handling, as well as other alien factors that they try to resist and avoid (16, 20). Stressful procedures and environmental factors are numerous and varied. Briefly, they include, but are not limited to: general handling and manipulations, such as weighing and saline injections (15, 16, 21–23); anaesthesia (15, 24–31); restraint (21, 32–36); gavage (37–42); blood sampling (15–17, 43–50); food and water restriction (51, 52); non-natural environment (53–55) and associated factors, such as noise and light (56–62); social crowding and/or isolation (63–71); cage conditions/cleaning/changing (18, 58, 72–78); transport (19, 58, 79–83); observing procedures on, and killing of, other animals (31, 84–87); and even

enrichment itself (17, 88, 89). It has been suggested that captive-bred animals may not know that these stressors are different to those in the wild, though there might be some perception that they are not similar to 'natural' stressors such as limited food, inadequate shelter, predation and so on. In any case, the point is that they are different, but more importantly, they are also frequent, regular and inescapable.

Naturally, this has animal welfare and scientific implications, acknowledged at least in some quarters. For instance, stress from handling is accepted as a source of "unexplained variation within and between animal studies", as it influences "both the behaviour and physiology of animals" (23; see also Balcombe *et al.* [16] and Meijer *et al.* [17]), and relatively poor caging conditions "may contribute to problems in translating murine research into human studies" (18, 77, 78). However, it is accepted by some that these factors are probably widely underappreciated (90). It should be noted that some consider enriched environments simply as 'less bad' rather than 'better' than those that are non-enriched. To illustrate, significant numbers of animals experiencing enrichment still go on to develop stereotypies (e.g. 91–98).

The Nature of Stress: Biological Basis/Mechanisms of Stress and its Adverse Effects

The physiological consequences of stress are varied and powerful. This, in itself, is of concern for the translation of animal data to humans, as they compound and confound existing difficulties in translation due to species differences. First, however, a consideration of the underlying mechanisms is important, to demonstrate their fundamental nature and potency.

Primary mediators of stress, such as GCs and catecholamines, are released in response to stressors, with various biological consequences. Such biological effects generally go beyond species boundaries/limits for mammalian species, though the mechanisms and specific effects differ to varying degrees. These primary mediators are extremely powerful, because they ultimately modulate the expression of many genes. Secondary outcomes have been documented "...in every physiological system, including the cardiovascular system, metabolism, the central nervous system, and the immune system", and are confounded by the characteristics of the stressor(s), as well as the attributes of the affected individual, such as age, health, status, genetic background, past experience, etc. (99).

The potency and ubiquity of the stress response has been demonstrated in a number of *in vitro* studies, revealing that it generally blocks every important cellular process, including DNA replication,

transcription, pre-mRNA processing, mRNA export, and translation, until the cells recover (100). Therefore, stress exerts its effects via varied molecular mechanisms, with far-reaching consequences. The principal ones are highlighted below.

- *Epigenetic mechanisms (histone acetylation and DNA methylation)* (101–107): Psychological stress alters gene expression via histone acetylation and DNA methylation. Much occurs in response to environmental triggers, e.g. diet, drugs, toxins, and psychological stress, e.g. fear conditioning and maternal care (103). Genes involved in HPA axis function are especially susceptible (108), e.g. in suicide victims with a history of child abuse (109), and post-traumatic stress disorder (PTSD) is strongly associated with the epigenetic modification of genes involved in immune function and inflammation (110).
- *Alternative splicing/expression of regulatory microRNAs*: Alternative splicing of gene transcripts and microRNAs (miRNAs), is a powerful means of altering gene expression that can be significantly affected by stress (111, 112). For example, acute stress in humans altered the splicing of 27 genes in peripheral leukocytes (100).
- *Oxidative damage and ageing*: Mental stress contributes to oxidative stress in the body, and therefore to oxidative damage (113). This effect has been identified in students undergoing academic examinations (114), and in the lymphocytes of psychologically stressed individuals (115). Oxidative stress is also associated with PTSD and depression (116), contributes to the ageing process (117), and is associated with neurodegenerative disease, ophthalmologic disease, cancer and cardiovascular disease (including atherosclerosis, hypertension, cardiomyopathy, chronic heart failure, myocardial ischaemia and ventricular arrhythmias; 117, 118). There may be confounding data on the effects of oxidative stress from different species/strains of mice, which further challenges the translation of data across species (119).

Direct Physiological Consequences of Stress

The physiological consequences of stress are numerous and varied. Table 1 shows how promiscuous these consequences, effects and manifestations are in many species, including humans. Given that these effects involve so many biological pathways and systems, the effects on experimental data must be significant.

Habituation/Desensitisation to Stress

Some argue that animals become habituated (120) and/or desensitised (121) to stress, so implications for welfare and experimental results may be overcome (23, 122, 123). For instance, non-human primates can be trained to approach test environments, and to present their arms for blood withdrawal and so on, seemingly voluntarily. However, it has been shown that repeat exposure to homotypic stressors of greater intensity and/or severity does not result in habituation, and could actually result in *sensitisation* of the CORT response (122). Furthermore, CORT levels might only decrease for certain types of stressor, persisting for other types (35); where desensitisation has been shown, it is only to a modest degree, in a small proportion of the animals in the studies, and in small sample sizes (70). While some studies have suggested that enrichment might decrease stress, others have shown a paradoxical increase in stress via CORT levels (18). Similarly, some studies have suggested that transferring scent-marked materials from old to new cages reduces stress-related aggression, while others found that it increases aggression (74). Furthermore, mice do not habituate to stress associated with simple handling, and indeed seem to become sensitised to it (124–126). In instances where CORT levels decrease, there is increasing evidence that this does not necessarily mean an attendant decrease in stress; other indicators, such as the neutrophil–lymphocyte ratio, might be ‘better’ indicators of chronic stress, which can occur in the absence of increased serum CORT (127).

Adverse Physiological Sequelae of Psychological Stress are Initiated Prenatally or in Early Life, and are Heritable

Captive-born animals can be exposed to stress prenatally via their wild-trapped mothers (128–131), and as infants in laboratory environments, often experiencing inadequate maternal contact and care (132–134; cited by Camus *et al.* [135], and Detter *et al.* [136]). If an animal’s parents or grandparents lived in laboratories, and/or were born of parents that lived in laboratories and/or endured being wild-caught, then their ancestors experienced highly stressful lives and would have been affected by the adverse consequences described herein. Even if such an individual was subsequently afforded as stress-free a life as possible (difficult, if not impossible, in a laboratory), the consequences of their early lives, and the lives of their ancestors, would lead to the same adverse effects as if they had continued to experience excessive stress as adults.

That early-life experiences affect adult psychopathology is widely accepted. As Jean-Paul Sartre put it, “Childhood decides” (see Murgatroyd [102]). “A large body of data shows that stress during pregnancy causes an increase of GCs in the blood of the dams and the foetus, leading to alterations of the structure and function of the developing brain... These alterations result in the disturbance of the function of the neuroendocrine system and different kinds of behaviour throughout life” (137). Early-life/prenatal exposure to stress leads to altered adrenocorticotrophic hormone responsiveness, dysfunction of the HPA axis (138, 139), and altered autonomic modulation of immune function that may begin *in utero* (103). Social isolation in several species leads to neuroendocrine changes, increased cortisol, and ensuing behavioural problems (for references, see Champagne [101]). Maternal inflammation during pregnancy (from infection, or possibly stress) may increase the risk of neurodevelopmental disorders such as schizophrenia and cerebral palsy (140). Physiological sequelae include cardiovascular disease and metabolic disorders such as diabetes (141), compromised immune function, including poor lymphocyte proliferation upon infection and reduced placental transfer of antibodies during pregnancy (140), autoimmune disorders, chronic obstructive lung disease, asthma and obesity (see Chang [142]). Pivotal to these adverse outcomes are the aforementioned stress-related epigenetic processes and oxidative damage. Methylation of gene regulatory regions is partly involved (143), the extent of which may be set during prenatal development (141). Cord blood samples of infants of mothers with late-pregnancy depression show altered methylation of the GC receptor promoter, which also predicts elevated salivary cortisol in early life (144). Other modifications are inherited and transgenerational in nature (145); for example, poor prenatal nutrition affects GC receptor methylation, affecting the growth and metabolism of first and second-generation offspring (146), and matrilineal transmission of the effects of diethylstilbestrol (DES) occurs via hypomethylation, leading to increased cancer risk over two generations (147).

Summary

The inherent, multi-faceted stress of laboratory life — often excessive, relative to the more transient, acute and ‘natural’ stresses experienced in the wild — is evidenced by its often negative impact on the well-being of the animals involved, both psychologically and physiologically. Because this harm is mediated via established trans-species biological mechanisms involving the HPA axis and the sympathetic nervous system, and effected via oxidative

stress and epigenetic mechanisms, which affect multiple biological pathways and systems, it can have adverse and confounding effects on experimental results. This modulation of many biochemical pathways and gene expression can result in downstream effects such as organ damage, cardiovascular diseases, attenuated immune function and autoimmune disorders, premature ageing and mortality, developmental abnormalities, elevated tumour initiation and progression, and musculoskeletal atrophy (16).

The absolute degree of translation of animal studies to humans is debatable, but undoubtedly, “Studies using animal models are more translatable to human disease when the animals’ welfare is maximised” (127). Arguably, it is difficult to “maximise” welfare significantly, given the inherent, widespread, substantial and largely intractable nature of the stresses involved in animal research and laboratory life. Notably, habituation and/or desensitisation to many of the stressors is often not possible, or at least not significant, and the impact of these effects on experimental data — and their extrapolation to humans — is likely to be significant. The many sources of stress have been summarised here, along with their effects on multiple biological and physiological systems. The literature warns that: “...animals subjected to the environmental changes that occur during transportation... react with changes in their physiology, such as body weight, plasma hormonal levels, heart rate and blood pressure changes... When measurements of physiological parameters are performed using conventional measurement techniques, the results must be interpreted with caution as these conventional techniques also have effects on the animals” (148). Most importantly, “Suffering in animals can result in physiological changes which may increase the variability of experimental data” (149). Many scientists are well aware of these effects and considerations, and have cautioned against disregarding them (150–152). Yet, while accepting the negative effects of pain, stress and distress, and their influence on study outcome, such effects are often not reported or are under-reported in scientific publications (90).

The impact of stress on immunological and inflammatory responses seems particularly prevalent, and might be especially critical, seeing as much animal experimentation involves infectious agents and/or immune function (for a discussion and references, see Bailey [153] and Bailey [154]). Crucially, this impact exacerbates and compounds existing immune differences between humans and non-humans due to genetic differences. To illustrate, genomic duplications — one of the most significant causes of genetic variation among primates (155) and at the root of many aspects of intra-species and inter-species diversity — differentially affect many genes involved in immune and

Table 1: A summary of the physiological consequences and manifestations of stress, in many species, including humans

Consequences of general stress	Notes
Stereotypes (abnormal, repetitive, invariant behaviours with no obvious function; 160, 161) and self-harm (e.g. 162–165) in non-humans	These correlate with basal plasma cortisol and corticotrophin-releasing hormone (CRH) in cerebrospinal fluid (166).
Wide-ranging physiological symptoms in humans	Anger, depression, anxiety, behavioural changes, food cravings, lack of appetite, frequent crying, difficulty sleeping, tiredness, lack of concentration, chest pains, constipation, diarrhoea, cramps and muscle spasms, dizziness, fainting, nervous twitches, restlessness, sexual dysfunctions, breathlessness, and a host of diseases and illnesses with probable associated psychogenic (as well as biological) causes (167).
Long-lasting neurophysiological changes	<p>These could “...have direct implications for electrophysiological, behavioural, and molecular studies” (see 69).</p> <ul style="list-style-type: none"> — Isolated rats show “...structural and functional changes in the mesocorticolimbic dopaminergic system, exhibited hyperlocomotor activity and impaired sensorimotor gating” (168). — Isolated pigs show “...sustained changes in behavioural, neuroendocrine and immune regulation” (169). — Socially isolated humans show increased risk of death, “...genome-wide transcriptional activity of impaired GC response genes and increased activity of pro-inflammatory transcription control pathways”, and higher risk of developing “...conduct disorders, personality disorders, major depression, PTSD, schizophrenia, and anxiety disorders” (see 170).
Multi-faceted modulation of the immune system	<p>This occurs in many, if not all, mammalian species (140, 171, 172). It is especially problematic for research involving immune function, infectious diseases, etc. The expression of several hundred genes (many related to immune function) may be perturbed by simple handling of animals (173; and see 33), which may be affected by the acute or chronic nature of the stressor (see 173). Chronic stress can:</p> <ul style="list-style-type: none"> — shift neutrophil-lymphocyte ratios (127, 174); — affect expression of cytokines and cytokine receptors (175, 176), such as IL-2 and IL-6 (18); — alter gene splicing in peripheral leukocytes (10); — alter global and immune gene specific methylation (177); and — differentially affect brain activity and neurotransmitter release, macrophage activity and antibody production (33). <p>The same stressor in acute or chronic forms may have different effects:</p> <ul style="list-style-type: none"> — In rodents, acute restraint increases delayed-type hypersensitivity (DTH) and leukocyte redeployment, but can increase or decrease them when chronic (33, 178). — In humans, chronic, though not acute, stress increases susceptibility to colds (179). <p>These inconsistencies show that stress is mediated not only by GCs (180), and that accounting for the effects of stress in animal research must be difficult, if not impossible.</p>
Observed alterations in human immune function	<p>These include:</p> <ul style="list-style-type: none"> — “attenuated responses to vaccination, poorer wound healing, exaggerated release of inflammatory mediators, & premature ageing of the immune system” (181); — increased plasma and CNS cytokine levels, impaired natural killer cell activity, lower T-lymphocyte counts (in PTSD/complex PTSD patients; 182); — epigenetic changes exerting lifelong impact on immune and inflammatory function (in PTSD/complex PTSD patients; 182); and — greater inflammatory responses to vaccinations (in depressed humans; 183). <p>Neuropeptides involved in stress responses may accentuate pathophysiological sequelae in critically ill individuals (184). In healthy humans, 49 different genetic pathways are affected by stress, including genes associated with immune function (185). Stressed students undergoing examinations have significantly increased pro-inflammatory cytokines (186).</p>
Activation of the HPA axis	<p>This accelerates ageing generally, with adverse effects on brain/central nervous system, immune system, skeletal muscle and bone tissue (see 187–189). HPA dysregulation may lead to excessive inflammation via increases in the levels of circulatory inflammatory cytokines, decreases in anti-inflammatory cytokines, and alterations in the expression of genes involved in immune activation of peripheral blood cells (see 177).</p>

Table 1: continued

Effects of specific stressors	Notes
Disease susceptibility	An increase in susceptibility has been noted across several species to: <ul style="list-style-type: none"> — general disease and somatic disorders (see 100, 103); — various cancers (190, 191); — pancreatitis and pancreatic tumours (192, 193); — gastrointestinal disorders (194); — thyroid pathology (195); — multiple sclerosis (171); — inflammatory bowel disease (142, 196); — cardiovascular disease (197–199); — accelerated ageing and age-related disorders (187); and — musculoskeletal injury (200). Stressors can increase oxidative stress, triggering inflammatory pathways associated with type-2 diabetes, cardiovascular disease, osteoporosis, arthritis, some cancers, and susceptibility to some infections (e.g. 201–205).
Handling	Causes biological changes, affecting wellbeing and/or experimental results. Enrichment intended to mitigate stress can substantially alter brain structure, function and physiology. Light, noise, cage position/changing etc. all affect physiology, behaviour, anxiety and experimental data (206). Effects of handling stress may often be “missed” by researchers (23).
Early-life stress in various species	Results in: <ul style="list-style-type: none"> — abnormal brain development, leading to early-life psychopathologies and adult chronic mental illnesses (e.g. 207, 208); — epigenetic modifications associated with depression and suicidal behaviour in later life (e.g. 102, 209–211); and — elevated morbidity and mortality from chronic diseases of ageing, including vascular disease, autoimmune disorders, and premature mortality (e.g. 141, 212). Stress from maternal deprivation has negative effects on growth rates in rats, and adversely affects circadian clock and stress responses (72).
Traumatic stress in humans	Studies on PTSD patients have shown that: <ul style="list-style-type: none"> — acute stress affects glucose metabolism, inflammation and components of the immune system associated with type-2 diabetes (201); — serious long-term consequences include hypertension, heart attacks and stroke, as well as increased risk of obesity, Alzheimer’s disease, and AIDS dementia complex (213).

inflammatory responses (156). Indels (genomic insertions and deletions) also affect major histocompatibility complex (MHC) genes, which are critical to immune responses, and are associated with differences in response to infections, as well as susceptibility to autoimmune diseases. These are further confounded by sex-related and strain-related differences. It has also been shown that the susceptibility and responsiveness of mice to stressors varies with the strain (157, 158). Stress also affects sleep, and conversely sleep perturbations exacerbate and sensitise individuals to stress, with all the attendant consequences. These consequences are also strain-dependent, and intimately linked to the CRH/HPA system (159).

Additionally, the adverse consequences of stress are multigenerational, as the associated epigenetic mechanisms affect the germline. This is likely to have significant consequences for animals, and their offspring, in laboratories: if an animal's parents or grandparents experienced a stressful laboratory life and experimental procedures, and/or if the offspring experienced significant stress in early life, then this will compound any further stress that they experience as adults, in turn compounding species differences and the translation of data to humans.

Overall, these observations of detrimental physiological effects and the general mechanisms behind them have been detailed in many species (including humans), and throughout the evolutionary scale from monkeys to rodents. The minutiae of the genes and biochemical pathways responsible, and their manifestations, may differ to some degree, but there are common mechanisms and adverse effects in all species examined to date. It must be concluded that laboratory life for animals used in experiments has serious and intractable consequences for their welfare, and for the quality and human relevance of the experimental data obtained (which, in any case, are already of debatable applicability to humans, due to species differences).

Finally, I believe that this issue should be taken much more seriously by legislators, regulators, funders, practitioners and advocates of animal experiments, and urge all involved to do so. The information presented here could, and should, be a valuable resource for project licence applicants, ethics committees and the Home Office Inspectorate, for use in experimental design and harm-benefit analyses, and to aid data interpretation. Though it argues that relatively little can be done to minimise many, if not all, stressors and stress, it could inform attempts to do so — as well as controlling variable factors and mitigating negative consequences, etc. The information could also be factored into existing guidance for the strategic planning of animal experiments, since stress impacts all areas of this planning, including study

objectives, species/strain selection, experimental procedures, analgesia, training of staff, and so on.

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References

1. Bailey, J. (2017). Does the stress inherent to laboratory life and experimentation on animals adversely affect research data. *ATLA* **45**, 299–301.
2. Anon. (2007). *Stress*. Saunders Comprehensive Veterinary Dictionary, 3rd edn. Available at: <http://medical-dictionary.thefreedictionary.com/Stress> (Accessed 19.09.18).
3. Ferdowsian, H.R. & Beck, N. (2011). Ethical and scientific considerations regarding animal testing and research. *PLoS One* **6**, e24059.
4. Rey, S., Huntingford, F.A., Boltaña, S., Vargas, R., Knowles, T.G. & Mackenzie, S. (2015). Fish can show emotional fever: Stress-induced hyperthermia in zebrafish. *Proceedings of the Biological Sciences B* **282**, 20152266.
5. Tran, S., Nowicki, M., Fulcher, N., Chatterjee, D. & Gerlai, R. (2016). Interaction between handling induced stress and anxiolytic effects of ethanol in zebrafish: A behavioral and neurochemical analysis. *Behavioural Brain Research* **298**, 278–285.
6. Rambo, C.L., Mocelin, R., Marcon, M., Villanova, D., Koakoski, G., de Abreu, M.S., Oliveira, T.A., Barcellos, L.J.G., Piatto, A.L. & Bonan, C.D. (2017). Gender differences in aggression and cortisol levels in zebrafish subjected to unpredictable chronic stress. *Physiology & Behavior* **171**, 50–54.
7. Fulcher, N., Tran, S., Shams, S., Chatterjee, D. & Gerlai, R. (2017). Neurochemical and behavioral responses to unpredictable chronic mild stress following developmental isolation: The zebrafish as a model for major depression. *Zebrafish* **14**, 23–34.
8. Zimmermann, F.F., Altenhofen, S., Kist, L.W., Leite, C.E., Bogo, M.R., Cognato, G.P. & Bonan, C.D. (2016). Unpredictable chronic stress alters adenosine metabolism in zebrafish brain. *Molecular Neurobiology* **53**, 2518–2528.
9. Maestripieri, D. & Hoffman, C.L. (2011). Chronic stress, allostatic load, and aging in nonhuman primates. *Development & Psychopathology* **23**, 1187–1195.
10. National Research Council (US) Committee on Recognition and Alleviation of Distress in Laboratory Animals (2008). *Recognition and Alleviation of Pain and Distress in Laboratory Animals*, 198pp. Washington, DC, USA: National Academies Press. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK4032/> (Accessed 13.09.17).
11. Moberg, G.P. & Mench, J.A. (eds) (2000). *The Biology of Animal Stress: Basic Principles and Implications for Animal Welfare*, 377pp. Wallingford, UK: CABI Publishing.

12. Tracey, K.J. (2009). Reflex control of immunity. *Nature Reviews Immunology* **9**, 418–428.
13. Sternberg, E.M. (2006). Neural regulation of innate immunity: A coordinated nonspecific host response to pathogens. *Nature Reviews Immunology* **6**, 318–328.
14. Glaser, R. & Kiecolt-Glaser, J.K. (2005). Stress-induced immune dysfunction: Implications for health. *Nature Reviews Immunology* **5**, 243–251.
15. Altholtz, L.Y., Fowler, K.A., Badura, L.L. & Kovacs, M.S. (2006). Comparison of the stress response in rats to repeated isoflurane or CO₂:O₂ anesthesia used for restraint during serial blood collection via the jugular vein. *Journal of the American Association for Laboratory Animal Science* **45**(3), 17–22.
16. Balcombe, J.P., Barnard, N.D. & Sandusky, C. (2004). Laboratory routines cause animal stress. *Contemporary Topics in Laboratory Animal Science* **43**, 42–51.
17. Meijer, M.K., Sommer, R., Spruijt, B.M., van Zutphen, L.F. & Baumans, V. (2007). Influence of environmental enrichment and handling on the acute stress response in individually housed mice. *Laboratory Animals* **41**, 161–173.
18. Gurfein, B.T., Stamm, A.W., Bacchetti, P., Dallman, M.F., Nadkarni, N.A., Milush, J.M., Touma, C., Palme, R., Di Borgo, C.P., Fromentin, G., Lowenthal-Hecht, R., Konsman, J.P., Acree, M., Premenko-Lanier, M., Darcel, N., Hecht, F.M. & Nixon, D.F. (2012). The calm mouse: An animal model of stress reduction. *Molecular Medicine* **18**, 606–617.
19. Obernier, J.A. & Baldwin, R.L. (2006). Establishing an appropriate period of acclimatization following transportation of laboratory animals. *ILAR Journal* **47**, 364–369.
20. Balcombe, J.P. (2006). Laboratory environments and rodents' behavioural needs: A review. *Laboratory Animals* **40**, 217–235.
21. Meijer, M.K., Spruijt, B.M., van Zutphen, L.F.M. & Baumans, V. (2006). Effect of restraint and injection methods on heart rate and body temperature in mice. *Laboratory Animals* **40**, 382–391.
22. Novak, M.A., Hamel, A.F., Kelly, B.J., Dettmer, A.M. & Meyer, J.S. (2013). Stress, the HPA axis, and nonhuman primate well-being: A review. *Applied Animal Behaviour Science* **143**, 135–149.
23. Gouveia, K. & Hurst, J.L. (2013). Reducing mouse anxiety during handling: Effect of experience with handling tunnels. *PLoS One* **8**, e66401.
24. Conlee, K.M., Stephens, M.L., Rowan, A.N. & King, L.A. (2005). Carbon dioxide for euthanasia: Concerns regarding pain and distress, with special reference to mice and rats. *Laboratory Animals* **39**, 137–161.
25. Wong, D., Makowska, I.J. & Weary, D.M. (2013). Rat aversion to isoflurane versus carbon dioxide. *Biology Letters* **9**, 20121000.
26. Moody, C.M. & Weary, D.M. (2014). Mouse aversion to isoflurane versus carbon dioxide gas. *Applied Animal Behaviour Science* **158**, 95–101.
27. Fyer, M.R., Uy, J., Martinez, J., Goetz, R., Klein, D.F., Fyer, A., Liebowitz, M.R. & Gorman, J. (1987). CO₂ challenge of patients with panic disorder. *American Journal of Psychiatry* **144**, 1080–1082.
28. Danneman, P.J., Stein, S. & Walshaw, S.O. (1997). Humane and practical implications of using carbon dioxide mixed with oxygen for anesthesia or euthanasia of rats. *Laboratory Animal Science* **47**, 376–385.
29. Anestis, S.F. (2009). Urinary cortisol responses to unusual events in captive chimpanzees (*Pan troglodytes*). *Stress* **12**, 49–57.
30. Whitten, P.L., Stavisky, R., Aureli, F. & Russell, E. (1998). Response of fecal cortisol to stress in captive chimpanzees (*Pan troglodytes*). *American Journal of Primatology* **44**, 57–69.
31. Springer, D.A. & Baker, K.C. (2007). Effect of ketamine anesthesia on daily food intake in *Macaca mulatta* and *Cercopithecus aethiops*. *American Journal of Primatology* **69**, 1080–1092.
32. Martin, C. (2014). Contributions and complexities from the use of *in vivo* animal models to improve understanding of human neuroimaging signals. *Frontiers in Neuroscience* **8**, 211.
33. Bowers, S.L., Bilbo, S.D., Dhabhar, F.S. & Nelson, R.J. (2008). Stressor-specific alterations in corticosterone and immune responses in mice. *Brain, Behavior, & Immunity* **22**, 105–113.
34. Maninger, N., Mason, W., Ruys, J., Mendoza, S. & Maninger, C. (2010). Acute and chronic stress increase DHEAS concentrations in rhesus monkeys. *Psychoneuroendocrinology* **35**, 1055–1062.
35. Lee, J.I., Shin, J.S., Lee, J.E., Jung, W.Y., Lee, G., Kim, M.S., Park, C.G. & Kim, S.J. (2013). Changes of N/L ratio and cortisol levels associated with experimental training in untrained rhesus macaques. *Journal of Medical Primatology* **42**, 10–14.
36. Reinhardt, V., Liss, C. & Stevens, C. (1995). Restraint methods of laboratory non-human primates: A critical review. *Animal Welfare* **4**, 221–238.
37. de Meijer, V.E., Le, H.D., Meisel, J.A. & Puder, M. (2010). Repetitive orogastric gavage affects the phenotype of diet-induced obese mice. *Physiology & Behavior* **100**, 387–393.
38. Ökva, K., Tamoseviciute, E., Ciziute, A., Pokk, P., Ruksenas, O. & Nevalainen, T. (2006). Refinements for intragastric gavage in rats. *Scandinavian Journal of Laboratory Animal Sciences* **33**, 243–252.
39. Gonzales, C., Zaleska, M.M., Riddell, D.R., Atchison, K.P., Robshaw, A., Zhou, H. & Sukoff Rizzo, S.J. (2014). Alternative method of oral administration by peanut butter pellet formulation results in target engagement of BACE1 and attenuation of gavage-induced stress responses in mice. *Pharmacology, Biochemistry & Behavior* **126**, 28–35.
40. Bonnichsen, M., Dragsted, N. & Hansen, A.K. (2005). The welfare impact of gavaging laboratory rats. *Animal Welfare* **14**, 223–227.
41. Brown, A.P., Dinger, N. & Levine, B.S. (2000). Stress produced by gavage administration in the rat. *Contemporary Topics in Laboratory Animal Science* **39**, 17–21.
42. Walker, M.K., Boberg, J.R., Walsh, M.T., Wolf, V., Trujillo, A., Duke, M.S., Palme, R. & Felton, L.A. (2012). A less stressful alternative to oral gavage for pharmacological and toxicological studies in mice. *Toxicology & Applied Pharmacology* **260**, 65–69.
43. Morton, D.B., Abbot, D., Barclay, R., Close, B.S., Ewbank, R., Gask, D., Heath, M., Mattic, S., Poole, T., Seamer, J., Southee, J., Thompson, A., Trussell, B., West, C. & Jennings, M. (1993). Removal of blood from laboratory mammals and birds. First report of the BVA/FRAME/RSPCA/UFAW Joint Working Group on Refinement. *Laboratory Animals* **27**, 1–22.
44. Sarlis, N.J. (1991). Chronic blood sampling techniques in stress experiments in the rat — A mini

- review. *Animal Technology* **42**, 51–59.
45. Capitanio, J.P., Mendoza, S.P. & McChesney, M. (1996). Influences of blood sampling procedures on basal hypothalamic-pituitary-adrenal hormone levels and leukocyte values in rhesus macaques (*Macaca mulatta*). *Journal of Medical Primatology* **25**, 26–33.
46. Teilmann, A.C., Kallikoski, O., Sorensen, D.B., Hau, J. & Abelson, K.S. (2014). Manual versus automated blood sampling: Impact of repeated blood sampling on stress parameters and behavior in male NMRI mice. *Laboratory Animals* **48**, 278–291.
47. Teilmann, A.C., Nygaard Madsen, A., Holst, B., Hau, J., Rozell, B. & Abelson, K.S. (2014). Physiological and pathological impact of blood sampling by retrobulbar sinus puncture and facial vein phlebotomy in laboratory mice. *PLoS One* **9**, e113225.
48. Burnett, J.E. (2011). Dried blood spot sampling: Practical considerations and recommendation for use with preclinical studies. *Bioanalysis* **3**, 1099–1107.
49. Wickremsinhe, E.R. & Perkins, E.J. (2015). Using dried blood spot sampling to improve data quality and reduce animal use in mouse pharmacokinetic studies. *Journal of the American Association for Laboratory Animal Science* **54(2)**, 139–144.
50. Langkilde, T. & Shine, R. (2006). How much stress do researchers inflict on their study animals? A case study using a scincid lizard, *Eulamprus heatwolei*. *Journal of Experimental Biology* **209**, 1035–1043.
51. Jennings, M., Prescott, M.J., Buchanan-Smith, H.M., Gamble, M.R., Gore, M., Hawkins, P., Hubrecht, R., Hudson, S., Jennings, M., Keeley, J.R., Morris, K., Morton, D.B., Owen, S., Pearce, P.C., Prescott, M.J., Robb, D., Rumble, R.J., Wolfensohn, S. & Buist, D. (2009). Refinements in husbandry, care and common procedures for non-human primates: Ninth report of the BVAWF/FRAME/RSPCA/UFAW Joint Working Group on Refinement. *Laboratory Animals* **43**, Suppl. 1, 1–47.
52. Willems, R.A. (2009). Regulatory issues regarding the use of food and water restriction in laboratory animals. *Lab Animal* **38**, 325–328.
53. Calisi, R.M. & Bentley, G.E. (2009). Lab and field experiments: Are they the same animal? *Hormones & Behavior* **56**, 1–10.
54. Meijer, T. & Schwabl, H. (1989). Hormonal patterns in breeding and nonbreeding kestrels, *Falco tinnunculus*: Field and laboratory studies. *General & Comparative Endocrinology* **74**, 148–160.
55. Marra, P.P., Kevin, T.L. & Bruce, L.T. (1995). Plasma corticosterone levels in two species of *Zonotrichia* sparrows under captive and free-living conditions. *Wilson Bulletin* **107**, 305–296.
56. Barrett, A.M. & Stockham, M.A. (1963). The effect of housing conditions and simple experimental procedures upon the corticosterone level in the plasma of rats. *Journal of Endocrinology* **26**, 97–105.
57. Cavigelli, S.A., Guhad, F.A., Ceballos, R.M., Whetzel, C.A., Nevalainen, T., Lang, C.M. & Klein, L.C. (2006). Fecal corticoid metabolites in aged male and female rats after husbandry-related disturbances in the colony room. *Journal of the American Association for Laboratory Animal Science* **45(6)**, 17–21.
58. Castelhano-Carlos, M.J. & Baumans, V. (2009). The impact of light, noise, cage cleaning and in-house transport on welfare and stress of laboratory rats. *Laboratory Animals* **43**, 311–327.
59. Baldwin, A.L. (2007). Effects of noise on rodent physiology. *International Journal of Comparative Psychology* **20**, 134–144.
60. Pines, M.K., Kaplan, G. & Rogers, L.J. (2004). Stressors of common marmosets (*Callithrix jacchus*) in the captive environment: Effects on behaviour and cortisol levels. *Folia Primatologica* **75**, 317–318.
61. Schreuder, M.F., Fodor, M., van Wijk, J.A. & Delemarre-van de Waal, H.A. (2007). Weekend versus working day: Differences in telemetric blood pressure in male Wistar rats. *Laboratory Animals* **41**, 86–91.
62. Patterson-Kane, E.G. & Farnworth, M.J. (2006). Noise exposure, music, and animals in the laboratory: A commentary based on Laboratory Animal Refinement and Enrichment Forum (LAREF) discussions. *Journal of Applied Animal Welfare Science* **9**, 327–332.
63. Kirillov, O.I., Khasina, E.I. & Durkina, V.B. (2003). [Effect of stress on postnatal growth in weight of rat body and adrenal gland]. *Ontogeny* **34**, 371–376.
64. Bernátová, I., Púzserová, A., Navarová, J., Csizmadiová, Z. & Zeman, M. (2007). Crowding-induced alterations in vascular system of Wistar-Kyoto rats: Role of nitric oxide. *Physiological Research* **56**, 667–669.
65. Armario, A., Castellanos, J.M. & Balasch, J. (1984). Effect of crowding on emotional reactivity in male rats. *Neuroendocrinology* **39**, 330–333.
66. Cyr, N.E. & Romero, L.M. (2008). Fecal glucocorticoid metabolites of experimentally stressed captive and free-living starlings: Implications for conservation research. *General & Comparative Endocrinology* **158**, 20–28.
67. Wingfield, J.C. & Kitaysky, A.S. (2002). Endocrine responses to unpredictable environmental events: Stress or anti-stress hormones. *Integrative & Comparative Biology* **42**, 600–609.
68. Romero, L.M. & Wingfield, J.C. (1999). Alterations in hypothalamic-pituitary-adrenal function associated with captivity in Gambel's white-crowned sparrows (*Zonotrichia leucophrys gambeli*). *Comparative Biochemistry & Physiology. Part B, Biochemistry & Molecular Biology* **122**, 13–20.
69. Ferland, C.L. & Schrader, L.A. (2011). Cage mate separation in pair-housed male rats evokes an acute stress corticosterone response. *Neuroscience Letters* **489**, 154–158.
70. Clay, A.W., Bloomsmith, M.A., Marr, M.J. & Maple, T.L. (2009). Habituation and desensitization as methods for reducing fearful behavior in singly housed rhesus macaques. *American Journal of Primatology* **71**, 30–39.
71. Lilly, A.A., Mehlman, P.T. & Higley, J.D. (1999). Trait-like immunological and hematological measures in female rhesus across varied environmental conditions. *American Journal of Primatology* **48**, 197–223.
72. Allen, K.P., Dwinell, M.R., Zappa, A., Temple, A. & Thulin, J. (2013). Comparison of 2 rat breeding schemes using conventional caging. *Journal of the American Association of Laboratory Animal Science* **52(2)**, 142–145.
73. Burn, C.C., Peters, A., Day, M.J. & Mason, G.J. (2006). Long-term effects of cage-cleaning frequency and bedding type on laboratory rat health, welfare, and handleability: A cross-laboratory study. *Laboratory Animals* **40**, 353–370.
74. Meller, A., Kasanen, I., Ruksenas, O., Apanaviciene,

- N., Baturaite, Z., Voipio, H.M. & Nevalainen, T. (2011). Refining cage change routines: Comparison of cardiovascular responses to three different ways of cage change in rats. *Laboratory Animals* **45**, 167–173.
75. Line, S.W., Markowitz, H., Morgan, K.N. & Strong, S. (1991). Effects of cage size and environmental enrichment on behavioral and physiological responses of rhesus macaques to the stress of daily events. In *Through the Looking Glass* (ed. M.A. Novak & A.J. Petto), pp. 160–179. Washington, DC, USA: American Psychological Association.
76. Clarke, A.S., Mason, W.A. & Mendoza, S.P. (1994). Heart rate patterns under stress in three species of macaques. *American Journal of Primatology* **33**, 133–148.
77. Würbel, H. (2001). Ideal homes? Housing effects on rodent brain and behaviour. *Trends in Neurosciences* **24**, 207–211.
78. Wolfer, D.P., Litvin, O., Morf, S., Nitsch, R.M., Lipp, H.P. & Würbel, H. (2004). Laboratory animal welfare: Cage enrichment and mouse behaviour. *Nature, London* **432**, 821–822.
79. Burn, C.C., Deacon, R.M. & Mason, G.J. (2008). Marked for life? Effects of early cage-cleaning frequency, delivery batch, and identification tail-marking on rat anxiety profiles. *Developmental Psychobiology* **50**, 266–277.
80. Schapiro, S.J., Lambeth, S.P., Jacobsen, K.R., Williams, L.E., Nehete, B.N. & Nehete, P.N. (2012). Physiological and welfare consequences of transport, relocation, and acclimatization of chimpanzees (*Pan troglodytes*). *Applied Animal Behaviour Science* **137**, 183–193.
81. Wolfensohn, S.E. (1997). Brief review of scientific studies of the welfare implications of transporting primates. *Laboratory Animals* **31**, 303–305.
82. Honess, P.E., Johnson, P.J. & Wolfensohn, S.E. (2004). A study of behavioural responses of non-human primates to air transport and re-housing. *Laboratory Animals* **38**, 119–132.
83. Ferreira, C.S., Vasconcellos, R.S., Pedreira, R.S., Silva, F.L., Sá, F.C., Kroll, F.S., Maria, A.P., Venturini, K.S. & Carciofi, A.C. (2014). Alterations to oxidative stress markers in dogs after a short-term stress during transport. *Journal of Nutritional Science* **3**, e27.
84. Line, S.W., Morgan, K.N., Markowitz, H. & Strong, S. (1989). Heart rate and activity of rhesus monkeys in response to routine events. *Laboratory Primate Newsletter* **28**, 9–12.
85. Sharp, J., Zammit, T., Azar, T. & Lawson, D. (2003). Are “by-stander” female Sprague-Dawley rats affected by experimental procedures. *Contemporary Topics in Laboratory Animal Science* **42**, 19–27.
86. Gilmore, A.J., Billing, R.L. & Einstein, R. (2008). The effects on heart rate and temperature of mice and vas deferens responses to noradrenaline when their cage mates are subjected to daily restraint stress. *Laboratory Animals* **42**, 140–148.
87. Bowers, C.L., Crockett, C.M. & Bowden, D.M. (1998). Differences in stress reactivity of laboratory macaques measured by heart period and respiratory sinus arrhythmia. *American Journal of Primatology* **45**, 245–261.
88. Baumans, V. (2005). Environmental enrichment for laboratory rodents and rabbits: Requirements of rodents, rabbits, and research. *ILAR Journal* **46**, 162–170.
89. Hutchinson, E., Avery, A. & Vandewoude, S. (2005). Environmental enrichment for laboratory rodents. *ILAR Journal* **46**, 148–161.
90. Reinhardt, V. & Reinhardt, A. (2000). Blood collection procedure of laboratory primates: A neglected variable in biomedical research. *Journal of Applied Animal Welfare Science* **3**, 321–333.
91. Olsson, A. & Dahlborn, K. (2002). Improving housing conditions for laboratory mice: A review of ‘environmental enrichment’. *Laboratory Animals* **36**, 243–270.
92. Zimmermann, A., Stauffacher, M., Langhans, W. & Würbel, H. (2001). Enrichment-dependent differences in novelty exploration in rats can be explained by habituation. *Behavioural Brain Research* **121**, 11–20.
93. Würbel, H., Chapman, R. & Rutland, C. (1998). Effect of feed and environmental enrichment on development of stereotypic wire-gnawing in laboratory mice. *Applied Animal Behaviour Science* **60**, 69–81.
94. Powell, S.B., Newman, H.A., McDonald, T.A., Bugenhagen, P. & Lewis, M.H. (2000). Development of spontaneous stereotyped behavior in deer mice: Effects of early and late exposure to a more complex environment. *Developmental Psychobiology* **37**, 100–108.
95. Callard, M.D., Bursten, S.N. & Price, E.O. (2000). Repetitive backflipping behaviour in captive roof rats (*Rattus rattus*) and the effects of cage enrichment. *Animal Welfare* **9**, 139–152.
96. Balcombe, J. (2010). Laboratory rodent welfare: Thinking outside the cage. *Journal of Applied Animal Welfare Science* **13**, 77–88.
97. Leach, M.C., Ambrose, N., Bowell, V.J. & Morton, D.B. (2000). The development of a novel form of mouse cage enrichment. *Journal of Applied Animal Welfare Science* **3**, 81–91.
98. Würbel, H., Freire, R. & Nicol, C.J. (1998). Prevention of stereotypic wire-gnawing in laboratory mice: Effects on behaviour and implications for stereotypy as a coping response. *Behavioural Processes* **42**, 61–72.
99. McEwen, B.S. & Seeman, T. (1999). Protective and damaging effects of mediators of stress. Elaborating and testing the concepts of allostasis and allostatic load. *Annals of the New York Academy of Sciences* **896**, 30–47.
100. Kurokawa, K., Kuwano, Y., Tominaga, K., Kawai, T., Katsuura, S., Yamagishi, N., Satake, Y., Kajita, K., Tanahashi, T. & Rokutan, K. (2010). Brief naturalistic stress induces an alternative splice variant of SMG-1 lacking exon 63 in peripheral leukocytes. *Neuroscience Letters* **484**, 128–132.
101. Champagne, F.A. (2010). Epigenetic influence of social experiences across the lifespan. *Developmental Psychobiology* **52**, 299–311.
102. Murgatroyd, C. & Spengler, D. (2011). Epigenetic programming of the HPA axis: Early life decides. *Stress* **14**, 581–589.
103. Wright, R. (2011). Epidemiology of stress and asthma: From constricting communities and fragile families to epigenetics. *Immunology & Allergy Clinics of North America* **31**, 19–39.
104. Hoffmann, A., Zimmermann, C.A. & Spengler, D. (2015). Molecular epigenetic switches in neurodevelopment in health and disease. *Frontiers in Behavioral Neuroscience* **9**, 120.
105. Grace, C.E., Kim, S.J. & Rogers, J.M. (2011). Mat-

- ernal influences on epigenetic programming of the developing hypothalamic–pituitary–adrenal axis. *Birth Defects Research. Part A, Clinical & Molecular Teratology* **91**, 797–805.
106. Gudsnuk, K. & Champagne, F.A. (2012). Epigenetic influence of stress and the social environment. *ILAR Journal* **53**, 279–288.
 107. Champagne, F.A. (2012). Interplay between social experiences and the genome: Epigenetic consequences for behavior. *Advances in Genetics* **77**, 33–57.
 108. Wright, R.J. & Enlow, M.B. (2008). Maternal stress and perinatal programming in the expression of atopy. *Expert Review of Clinical Immunology* **4**, 535–538.
 109. McGowan, P.O., Sasaki, A., D'Alessio, A.C., Dymov, S., Labonte, B., Szyf, M., Turecki, G. & Meaney, M.J. (2009). Epigenetic regulation of the glucocorticoid receptor in human brain associates with childhood abuse. *Nature Neuroscience* **12**, 342–348.
 110. Uddin, M., Aiello, A.E., Wildman, D.E., Koenen, K.C., Pawelec, G., de los Santos, R., Goldmann, E. & Galea, S. (2010). Epigenetic and immune function profiles associated with posttraumatic stress disorder. *Proceedings of the National Academy of Sciences of the USA* **107**, 9470–9475.
 111. Biamonti, G. & Caceres, J.F. (2009). Cellular stress and RNA splicing. *Trends in Biochemical Sciences* **34**, 146–153.
 112. Katsura, S., Kuwano, Y., Yamagishi, N., Kurokawa, K., Kajita, K., Akaike, Y., Nishida, K., Masuda, K., Tanahashi, T. & Rokutan, K. (2012). MicroRNAs miR-144/144* and miR-16 in peripheral blood are potential biomarkers for naturalistic stress in healthy Japanese medical students. *Neuroscience Letters* **516**, 79–84.
 113. Hapuarachchi, J.R., Chalmers, A.H., Winefield, A.H. & Blake-Mortimer, J.S. (2003). Changes in clinically relevant metabolites with psychological stress parameters. *Behavioral Medicine* **29**, 52–59.
 114. Sivonova, M., Zitnanova, I., Hlincikova, L., Skodacek, I., Trebaticka, J. & Durackova, Z. (2004). Oxidative stress in university students during examinations. *Stress* **7**, 183–188.
 115. Knickelbein, K.Z., Flint, M., Jenkins, F. & Baum, A. (2008). Psychological stress and oxidative damage in lymphocytes of aerobically fit and unfit individuals. *Journal of Applied Biobehavioral Research* **13**, 1–19.
 116. Maes, M. (2001). Psychological stress and the inflammatory response system. *Clinical Science* **101**, 193–194.
 117. Videan, E.N., Heward, C.B., Chowdhury, K., Plummer, J., Su, Y. & Cutler, R.G. (2009). Comparison of biomarkers of oxidative stress and cardiovascular disease in humans and chimpanzees (*Pan troglodytes*). *Comparative Medicine* **59**, 287–296.
 118. Wang, L., Muxin, G., Nishida, H., Shirakawa, C., Sato, S. & Konishi, T. (2007). Psychological stress-induced oxidative stress as a model of sub-healthy condition and the effect of TCM. *Evidence-Based Complementary & Alternative Medicine* **4**, 195–202.
 119. Lewis, K.N., Andziak, B., Yang, T. & Buffenstein, R. (2013). The naked mole-rat response to oxidative stress: Just deal with it. *Antioxidants & Redox Signaling* **19**, 1388–1399.
 120. Webster, J. (2002). *Animal Welfare: A Cool Eye Towards Eden*, 284pp. Oxford, UK: Blackwell Science.
 121. Chance, P. (2003). *Learning and Behavior* (5th Edition). Belmont, CA, USA: Thomson Wadsworth.
 122. Barnum, C.J., Blandino, P.J. & Deak, T. (2007). Adaptation in the corticosterone and hyperthermic responses to stress following repeated stressor exposure. *Journal of Neuroendocrinology* **19**, 632–642.
 123. Gruen, M.E., Thomson, A.E., Clary, G.P., Hamilton, A.K., Hudson, L.C., Meeker, R.B. & Sherman, B.L. (2013). Conditioning laboratory cats to handling and transport. *Lab Animal* **42**, 385–389.
 124. Longordo, F., Fan, J., Steimer, T., Kopp, C. & Luthi, A. (2011). Do mice habituate to “gentle handling?” A comparison of resting behavior, corticosterone levels and synaptic function in handled and undisturbed C57BL/6J mice. *Sleep* **34**, 679–681.
 125. Kramer, K., van de Weerd, H., Mulder, A., Van Heijningen, C., Baumans, V., Remie, R., Voss, H.P. & van Zutphen, B.F. (2004). Effect of conditioning on the increase of heart rate and body temperature provoked by handling in the mouse. *ATLA* **32**, Suppl. 1A, 177–181.
 126. Clement, J.G., Mills, P. & Brockway, B. (1989). Use of telemetry to record body temperature and activity in mice. *Journal of Pharmacological Methods* **21**, 129–140.
 127. Swan, M.P. & Hickman, D.L. (2014). Evaluation of the neutrophil–lymphocyte ratio as a measure of distress in rats. *Lab Animal* **43**, 276–282.
 128. Anon. (2009). *The Revision of the UK Directive on the Protection of Animals Used for Scientific Purposes, Volume II: Evidence*, 252pp. London, UK: The Stationery Office. Available at: <http://www.publications.parliament.uk/pa/ld200809/ldselect/ldeucom/164/164iii.pdf> (Accessed 14.09.18).
 129. Anon. (2006). *Action to be Taken Against Laos, Vietnam and Cambodia Monkey Business*. London, UK: Cruelty Free International. Available at: <https://www.crueltyfreeinternational.org/breaking-news/action-be-taken-against-laos-vietnam-and-cambodia-monkey-business> (Accessed 19.09.18).
 130. Anon. (2008). *Cambodia: The Trade in Primates for Research*. London, UK: Cruelty Free International.
 131. Anon. (2010). *Mauritius: The Trade in Primates for Research*. London, UK: Cruelty Free International.
 132. Suomi, S.J. (1991). Primate separation models of affective disorders. In *Neurobiology of Learning, Emotion and Affect* (ed. J. Madden), pp. 195–214. New York, NY, USA: Lippincott Williams and Wilkins.
 133. Nakamichi, M., Cho, F. & Minami, T. (1990). Mother–infant interactions of wild-born, individually-caged cynomolgus monkeys (*Macaca fascicularis*) during the first 14 weeks of infant life. *Primates* **31**, 213–224.
 134. Maestripieri, D., Martel, F.L., Nevison, C.M., Simpson, M.J. & Keverne, E.B. (1991). Anxiety in rhesus monkey infants in relation to interactions with their mother and other social companions. *Developmental Psychobiology* **24**, 571–581.
 135. Camus, S.M., Rochais, C., Blois-Heulin, C., Li, Q., Hausberger, M. & Bezard, E. (2013). Birth origin differentially affects depressive-like behaviours: Are captive-born cynomolgus monkeys more vulnerable to depression than their wild-born counterparts? *PLoS One* **8**, e67711.
 136. Dettmer, A.M., Novak, M.A., Suomi, S.J. & Meyer, J.S. (2012). Physiological and behavioral adaptation to relocation stress in differentially reared rhe-

- sus monkeys: Hair cortisol as a biomarker for anxiety-related responses. *Psychoneuroendocrinology* **37**, 191–199.
137. Butkevich, I., Mikhailenko, V., Semionov, P., Bagaeva, T., Otellin, V. & Aloisi, A.M. (2009). Effects of maternal corticosterone and stress on behavioral and hormonal indices of formalin pain in male and female offspring of different ages. *Hormones & Behavior* **55**, 149–157.
138. Heim, C., Newport, D.J., Wagner, D., Wilcox, M.M., Miller, A.H. & Nemeroff, C.B. (2002). The role of early adverse experience and adulthood stress in the prediction of neuroendocrine stress reactivity in women: A multiple regression analysis. *Depression & Anxiety* **15**, 117–125.
139. Heim, C., Mletzko, T., Purselle, D., Musselman, D.L. & Nemeroff, C.B. (2008). The dexamethasone/corticotropin-releasing factor test in men with major depression: Role of childhood trauma. *Biological Psychiatry* **63**, 398–405.
140. Christian, L.M. (2012). Psychoneuroimmunology in pregnancy: Immune pathways linking stress with maternal health, adverse birth outcomes, and fetal development. *Neuroscience & Biobehavioral Reviews* **36**, 350–361.
141. Kinnally, E.L., Feinberg, C., Kim, D., Ferguson, K., Leibel, R., Coplan, J.D. & John, M.J. (2011). DNA methylation as a risk factor in the effects of early life stress. *Brain, Behavior, & Immunity* **25**, 1548–1553.
142. Chang, L. (2011). The role of stress on physiologic responses and clinical symptoms in irritable bowel syndrome. *Gastroenterology* **140**, 761–765.
143. Bird, A.P. (1986). CpG-rich islands and the function of DNA methylation. *Nature, London* **321**, 209–213.
144. Oberlander, T.F., Weinberg, J., Papsdorf, M., Grunau, R., Misri, S. & Devlin, A.M. (2008). Prenatal exposure to maternal depression, neonatal methylation of human glucocorticoid receptor gene (NR3C1) and infant cortisol stress responses. *Epigenetics* **3**, 97–106.
145. Fukuda, S. & Taga, T. (2005). Cell fate determination regulated by a transcriptional signal network in the developing mouse brain. *Anatomical Science International* **80**, 12–18.
146. Zambrano, E., Martinez-Samayoa, P.M., Bautista, C.J., Deas, M., Guillen, L., Rodriguez-Gonzalez, G.L., Guzman, C., Larrea, F. & Nathanielsz, P.W. (2005). Sex differences in transgenerational alterations of growth and metabolism in progeny (F2) of female offspring (F1) of rats fed a low protein diet during pregnancy and lactation. *Journal of Physiology* **566**, 225–236.
147. Newbold, R.R., Padilla-Banks, E. & Jefferson, W.N. (2006). Adverse effects of the model environmental estrogen diethylstilbestrol are transmitted to subsequent generations. *Endocrinology* **147**, S11–S17.
148. Capdevila, S., Giral, M., Ruiz de la Torre, J.L., Russell, R.J. & Kramer, K. (2007). Acclimatization of rats after ground transportation to a new animal facility. *Laboratory Animals* **41**, 255–261.
149. Anon. (2007). NC3Rs blood sampling microsite launched. *Laboratory Animals* **41**, 407.
150. Mason, J.W., Wool, M.S., Wherry, F.E., Pennington, L.L., Brady, J.V. & Beer, B. (1968). Plasma growth hormone response to avoidance sessions in the monkey. *Psychosomatic Medicine* **30**, Suppl., 760–773.
151. Roberts, R.A., Soames, A.R., James, N.H., Gill, J.H. & Wheeldon, E.B. (1995). Dosing-induced stress causes hepatocyte apoptosis in rats primed by the rodent nongenotoxic hepatocarcinogen cyproterone acetate. *Toxicology & Applied Pharmacology* **135**, 192–199.
152. Brenner, G.J., Cohen, N., Ader, R. & Moynihan, J.A. (1990). Increased pulmonary metastases and natural killer cell activity in mice following handling. *Life Sciences* **47**, 1813–1819.
153. Bailey, J. (2011). Lessons from chimpanzee-based research on human disease: The implications of genetic differences. *ATLA* **39**, 527–540.
154. Bailey, J. (2014). Monkey-based research on human disease: The implications of genetic differences. *ATLA* **42**, 287–317.
155. Armengol, G., Knuutila, S., Lozano, J.J., Madrigal, I. & Caballin, M.R. (2010). Identification of human specific gene duplications relative to other primates by array CGH and quantitative PCR. *Genomics* **95**, 203–209.
156. Perry, G.H., Yang, F., Marques-Bonet, T., Murphy, C., Fitzgerald, T., Lee, A.S., Hyland, C., Stone, A.C., Hurles, M.E., Tyler-Smith, C., Eichler, E.E., Carter, N.P., Lee, C. & Redon, R. (2008). Copy number variation and evolution in humans and chimpanzees. *Genome Research* **18**, 1698–1710.
157. Tang, X., Orchard, S.M. & Sanford, L.D. (2002). Home cage activity and behavioral performance in inbred and hybrid mice. *Behavioural Brain Research* **136**, 555–569.
158. Tang, X. & Sanford, L.D. (2002). Telemetric recording of sleep and home cage activity in mice. *Sleep* **25**, 691–699.
159. Sanford, L.D., Yang, L., Wellman, L.L., Dong, E. & Tang, X. (2008). Mouse strain differences in the effects of corticotropin releasing hormone (CRH) on sleep and wakefulness. *Brain Research* **1190**, 94–104.
160. Mason, G.J. (1991). Stereotypies and suffering. *Behavioural Processes* **25**, 103–115.
161. Prescott, M.J., Morton, D.B., Anderson, D., Buckwell, A., Heath, M.S., Hubrecht, R., Jennings, M., Robb, M.D., Ruane, M.B. & Swallow, M.J. (2004). Refining dog husbandry and care. *Laboratory Animals* **38**, 1–94.
162. Hubrecht, R. (1995). The welfare of dogs in human care. In *The Domestic Dog: Its Evolution, Behaviour, and Interactions with People* (ed. J. Serpell), pp. 179–198. Cambridge, UK: Cambridge University Press.
163. Bourgeois, S.R., Vazquez, M. & Brasky, K. (2007). Combination therapy reduces self-injurious behavior in a chimpanzee (*Pan troglodytes troglodytes*): A case report. *Journal of Applied Animal Welfare Science* **10**, 123–140.
164. Brüne, M., Brüne-Cohrs, U., McGrew, W.C. & Preuschoft, S. (2006). Psychopathology in great apes: Concepts, treatment options and possible homologies to human psychiatric disorders. *Neuroscience & Biobehavioral Reviews* **30**, 1246–1259.
165. Lutz, C., Well, A. & Novak, M. (2003). Stereotypic and self-injurious behavior in rhesus macaques: A survey and retrospective analysis of environment and early experience. *American Journal of Primatology* **60**, 1–15.
166. Novak, M.A., Meyer, J.S., Lutz, C. & S., T. (2007). Stress and the performance of primate stereotypies. In *Stereotypic Animal Behaviour: Fundamentals and Applications for Welfare* (ed. G. Mason &

- J. Rushen), p. 248. Wallingford, UK: CAB International.
167. Anon. (2009). *Stress*. [Nursing Times, 23.02.09]. Available at: <https://www.nursingtimes.net/stress/1995960.article> (Accessed 17.09.18).
168. Wang, Y.C., Ho, U.C., Ko, M.C., Liao, C.C. & Lee, L.J. (2012). Differential neuronal changes in medial prefrontal cortex, basolateral amygdala and nucleus accumbens after postweaning social isolation. *Brain Structure & Function* **217**, 337–351.
169. Tuchscherer, M., Kanitz, E., Puppe, B., Tuchscherer, A. & Viergutz, T. (2009). Changes in endocrine and immune responses of neonatal pigs exposed to a psychosocial stressor. *Research in Veterinary Science* **87**, 380–388.
170. Kaushal, N., Nair, D., Gozal, D. & Ramesh, V. (2012). Socially isolated mice exhibit a blunted homeostatic sleep response to acute sleep deprivation compared to socially paired mice. *Brain Research* **1454**, 65–79.
171. Sorenson, M., Janusek, L. & Mathews, H. (2011). Psychological stress and cytokine production in multiple sclerosis: Correlation with disease symptomatology. *Biological Research for Nursing* **15**, 226–233.
172. Dhabhar, F.S. (2009). Enhancing versus suppressive effects of stress on immune function: Implications for immunoprotection and immunopathology. *Neuroimmunomodulation* **16**, 300–317.
173. He, Y.D., Karbowski, C.M., Werner, J., Everds, N., Di Palma, C., Chen, Y., Higgins-Garn, M., Tran, S., Afshari, C.A. & Hamadeh, H.K. (2014). Common handling procedures conducted in preclinical safety studies result in minimal hepatic gene expression changes in Sprague-Dawley rats. *PLoS One* **9**, e88750.
174. Li, H., Chen, L., Zhang, Y., Lesage, G., Zhang, Y., Wu, Y., Hanley, G., Sun, S. & Yin, D. (2011). Chronic stress promotes lymphocyte reduction through TLR2 mediated PI3K signaling in a beta-arrestin 2 dependent manner. *Journal of Neuroimmunology* **233**, 73–79.
175. Xiang, L., Rehm, K., Marshall, G. & Xiang, D.B. (2011). Effects of acute stress-induced immunomodulation on Th1/Th2 cytokine and catecholamine receptor expression in human peripheral blood cells. *Neuropsychobiology* **65**, 12–19.
176. Huang, C.J., Stewart, J.K., Franco, R.L., Evans, R.K., Lee, Z.P., Cruz, T.D., Webb, H.E. & Acevedo, E.O. (2011). LPS-stimulated tumor necrosis factor-alpha and interleukin-6 mRNA and cytokine responses following acute psychological stress. *Psychoneuroendocrinology* **36**, 1553–1561.
177. Smith, A., Conneely, K., Kilaru, V., Weiss, T., Bradley, B., Cubells, J., Ressler, K., Kilaru, M. & Bradley, T. (2011). Differential immune system DNA methylation and cytokine regulation in post-traumatic stress disorder. *American Journal of Medical Genetics. Part B, Neuropsychiatric Genetics* **156**, 700–708.
178. Dhabhar, F.S. & McEwen, B.S. (1997). Acute stress enhances while chronic stress suppresses cell-mediated immunity *in vivo*: A potential role for leukocyte trafficking. *Brain, Behavior, & Immunity* **11**, 286–306.
179. Cohen, S., Frank, E., Doyle, W.J., Skoner, D.P., Rabin, B.S. & Gwaltney, J.M. (1998). Types of stressors that increase susceptibility to the common cold in healthy adults. *Health Psychology* **17**, 214–223.
180. Blecha, F., Kelley, K.W. & Satterlee, D.G. (1982). Adrenal involvement in the expression of delayed-type hypersensitivity to SRBC and contact sensitivity to DNFB in stressed mice. *Proceedings of the Society for Experimental Biology & Medicine* **169**, 247–252.
181. Gouin, J.P., Hantsoo, L. & Kiecolt-Glaser, J.K. (2008). Immune dysregulation and chronic stress among older adults: A review. *Neuroimmunomodulation* **15**, 251–259.
182. Pace, T.W. & Heim, C.M. (2011). A short review on the psychoneuroimmunology of posttraumatic stress disorder: From risk factors to medical comorbidities. *Brain, Behavior, & Immunity* **25**, 6–13.
183. Glaser, R., Robles, T.F., Sheridan, J., Malarkey, W.B. & Kiecolt-Glaser, J.K. (2003). Mild depressive symptoms are associated with amplified and prolonged inflammatory responses after influenza virus vaccination in older adults. *Archive of General Psychiatry* **60**, 1009–1014.
184. Papathanassoglou, E.D., Giannakopoulou, M., Mpouzika, M., Bozas, E. & Karabinis, A. (2010). Potential effects of stress in critical illness through the role of stress neuropeptides. *Nursing in Critical Care* **15**, 204–216.
185. Nater, U.M., Whistler, T., Lonergan, W., Mletzko, T., Vernon, S.D. & Heim, C. (2009). Impact of acute psychosocial stress on peripheral blood gene expression pathways in healthy men. *Biological Psychology* **82**, 125–132.
186. Kamezaki, Y., Katsuura, S., Kuwano, Y., Tanahashi, T. & Rokutan, K. (2012). Circulating cytokine signatures in healthy medical students exposed to academic examination stress. *Psychophysiology* **49**, 991–997.
187. Hasan, K.M., Rahman, M.S., Arif, K.M. & Sobhani, M.E. (2011). Psychological stress and aging: Role of glucocorticoids (GCs). *Age* **34**, 1421–1433.
188. Kitajima, T., Ariizumi, K., Bergstresser, P.R. & Takashima, A. (1996). A novel mechanism of glucocorticoid-induced immune suppression: The inhibitor of T cell-mediated terminal maturation of a murine dendritic cell line. *Journal of Clinical Investigation* **98**, 142–147.
189. Porter, N.M. & Landfield, P.W. (1998). Stress hormones and brain aging: Adding injury to insult? *Nature Neuroscience* **1**, 3–4.
190. Sun, X., Zhong, X., Liu, Z., Cai, H., Fan, Q., Wang, Q., Liu, Q., Song, Y., He, S., Zhang, X. & Lu, Z. (2010). Proteomic analysis of chronic restraint stress-induced Gan (肝)-stagnancy syndrome in rats. *Chinese Journal of Integrative Medicine* **16**, 510–517.
191. Gidron, Y. & De, Z. (2010). Influence of stress and health-behaviour on miRNA expression. *Molecular Medicine Reports* **3**, 455–457.
192. Binker, M.G., Binker-Cosen, A.A., Richards, D., Gaisano, H.Y., de Cosen, R.H. & Cosen-Binker, L.I. (2010). Chronic stress sensitizes rats to pancreatitis induced by cerulein: Role of TNF- α . *World Journal of Gastroenterology* **16**, 5565–5581.
193. Schuller, H.M., Al-Wadei, H.A., Ullah, M.F. & Plummer, H.K. (2012). Regulation of pancreatic cancer by neuropsychological stress responses: A novel target for intervention. *Carcinogenesis* **33**, 191–196.
194. Hong, S., Owyang, C., Hong, Z. & Owyang, W. (2011). Corticosterone mediates reciprocal changes in CB1 and TRPV1 receptors in primary sensory neurons in the chronically stressed rat. *Gastro-*

- enterology **140**, 627–637.
195. Nadol'nik, L.I. (2010). [Stress and the thyroid gland]. *Biomeditsinskaia Khimiia* **56**, 443–456.
196. Rampton, D. (2011). The influence of stress on the development and severity of immune-mediated diseases. *Journal of Rheumatology Supplement* **88**, 43–47.
197. Menezes, A., Lavie, C., Milani, R., O'Keefe, J. & O'Keefe, L. (2011). Psychological risk factors and cardiovascular disease: Is it all in your head? *Postgraduate Medicine* **123**, 165–176.
198. Gavrilovic, L., Spasojevic, N. & Dronjak, S. (2010). Subsequent stress increases gene expression of catecholamine synthetic enzymes in cardiac ventricles of chronic-stressed rats. *Endocrine* **37**, 425–429.
199. Eitel, I., von Knobelsdorff-Brenkenhoff, F., Bernhardt, P., Carbone, I., Muellerleile, K., Aldrovandi, A., Francone, M., Desch, S., Gutberlet, M., Strohm, O., Schuler, G., Schulz-Menger, J., Thiele, H. & Friedrich, M.G. (2011). Clinical characteristics and cardiovascular magnetic resonance findings in stress (takotsubo) cardiomyopathy. *JAMA* **306**, 277–286.
200. Allen, D., McCall, G., Loh, A., Madden, M. & Mehan, R. (2010). Acute daily psychological stress causes increased atrophic gene expression and myostatin-dependent muscle atrophy. *American Journal of Physiology. Regulatory, Integrative & Comparative Physiology* **299**, R889–R898.
201. Nowotny, B., Cavka, M., Herder, C., Löffler, H., Poschen, U., Joksimovic, L., Kempf, K., Krug, A.W., Koenig, W., Martin, S. & Kruse, J. (2010). Effects of acute psychological stress on glucose metabolism and subclinical inflammation in patients with post-traumatic stress disorder. *Hormone & Metabolic Research* **42**, 746–753.
202. Marotta, F., Naito, Y., Padrini, F., Xuewei, X., Jain, S., Soresi, V., Zhou, L., Catanzaro, R., Zhong, K., Polimeni, A. & Chui, D.H. (2011). Redox balance signalling in occupational stress: Modification by nutraceutical intervention. *Journal of Biological Regulators & Homeostatic Agents* **25**, 221–229.
203. Jankord, R., Zhang, R., Flak, J.N., Solomon, M.B., Albertz, J. & Herman, J.P. (2010). Stress activation of IL-6 neurons in the hypothalamus. *American Journal of Physiology. Regulatory, Integrative & Comparative Physiology* **299**, R343–R351.
204. Videan, E.N., Fritz, J. & Murphy, J. (2008). Effects of aging on hematology and serum clinical chemistry in chimpanzees (*Pan troglodytes*). *American Journal of Primatology* **70**, 327–338.
205. Lammey, M., Baskin, G., Gigliotti, A., Lee, D.R., Ely, J. & Sleeper, M. (2008). Interstitial myocardial fibrosis in a captive chimpanzee (*Pan troglodytes*) population. *Comparative Medicine* **58**, 389–394.
206. Rasmussen, S., Miller, M.M., Filipski, S.B. & Tolwani, R.J. (2011). Cage change influences serum corticosterone and anxiety-like behaviors in the mouse. *Journal of the American Association for Laboratory Animal Science* **50(4)**, 479–483.
207. Wei, L., Simen, A., Mane, S. & Kaffman, A. (2012). Early life stress inhibits expression of a novel innate immune pathway in the developing hippocampus. *Neuropsychopharmacology* **37**, 567–580.
208. Jackowski, A., Perera, T.D., Abdallah, C.G., Garrido, G., Tang, C.Y., Martinez, J., Mathew, S.J., Gorman, J.M., Rosenblum, L.A., Smith, E.L., Dwork, A.J., Shungu, D.C., Kaffman, A., Gelernter, J., Coplan, J.D. & Kaufman, J. (2011). Early-life stress, corpus callosum development, hippocampal volumetrics, and anxious behavior in male nonhuman primates. *Psychiatry Research* **192**, 37–44.
209. Mehta, M. & Schmauss, C. (2011). Strain-specific cognitive deficits in adult mice exposed to early life stress. *Behavioral Neuroscience* **125**, 29–36.
210. Labonte, B. & Turecki, G. (2010). The epigenetics of suicide: Explaining the biological effects of early life environmental adversity. *Archives of Suicide Research* **14**, 291–310.
211. Schroeder, M., Krebs, M.O., Bleich, S. & Frieling, H. (2010). Epigenetics and depression: Current challenges and new therapeutic options. *Current Opinion in Psychiatry* **23**, 588–592.
212. Miller, G., Chen, E. & Chen, P. (2011). Psychological stress in childhood and susceptibility to the chronic diseases of aging: Moving toward a model of behavioral and biological mechanisms. *Psychological Bulletin* **137**, 959–997.
213. Raber, J. (1998). Detrimental effects of chronic hypothalamic–pituitary–adrenal axis activation. From obesity to memory deficits. *Molecular Neurobiology* **18**, 1–22.